



Oral Intake of Semi-refined Carrageenan by Rats Affects Apoptosis of Lymphocytes

Anton S. Tkachenko^{1,2*}, PhD;¹ Anatolii Onishchenko^{1,2}, PhD

¹Research Institute of Experimental and Clinical Medicine, Kharkiv National Medical University; Trinklera st. 6, 61022, Kharkiv, Ukraine

²Department of Biochemistry, Kharkiv National Medical University; Nauky av. 4, 61022, Kharkiv, Ukraine

*Corresponding authors: Anton S. Tkachenko, PhD; Ag. Director of the Research Institute of Experimental and Clinical Medicine, Kharkiv National Medical University, Nauky ave 4, 61022, Kharkiv, Ukraine. Tel: +380-50-1094554; Fax: +380-57-7004132; Email: antontkachenko555@gmail.com **Received:** 19-10-2020 **Revised:** 09-11-2020 **Accepted:** 24-12-2020



Introduction: The safety of generally recognized as safe food additives E407 and E407a is under rigorous debate. Our research aimed to evaluate the effects of the orally administered E407a (semi-refined carrageenan) on the viability of lymphocytes and their cell death modes.

Methods: In total, 16 adult WAG rats were divided into two equal groups (experimental – oral intake of 140 mg/kg of E407a for two weeks; control – oral consumption of drinking water instead). Blood samples were used to obtain leukocyte suspensions stained with Annexin V-FITC and 7-aminoactinomycin D (7-AAD). The region of lymphocytes was analyzed after collecting data using a BD FACSCanto[™] II flow cytometer.

Results: Oral administration of E407a led to a decrease in the number of viable (Annexin V⁻, 7-AAD⁻) circulating lymphocytes. Furthermore, exposure to semi-refined carrageenan resulted in an elevated number of early apoptotic (Annexin V⁺, 7-AAD⁻) lymphocytes; their percentage was approximately four times higher in rats exposed to E407a compared with the control group.

Conclusion: Our findings indicate that dietary intake of E407a promotes apoptosis of circulating lymphocytes.

Keywords: Processed Eucheuma seaweed, Annexin V, 7-aminoactinomycin D, Flow cytometry

Please cite this paper as:

Tkachenko AS, Onishchenko A. Oral Intake of Semi-refined Carrageenan by Rats Affects Apoptosis of Lymphocytes. *Ann Colorectal Res.* 2020;8(4):170-174. doi: 10.30476/ACRR.2021.88624.1068.

Introduction

Colorectal cancer (CRC) is known to be the second leading cancer-related cause of death (1). Despite some recent advances in its treatment, CRC remains a serious medical and social burden. According to some estimates, CRC was responsible for approximately 2 million deaths globally in 2018 (2). CRC is a heterogeneous disease with numerous modifiable factors, including dietary habits, physical activity, obesity, cigarette smoking, etc., which can contribute to its etiology (3). Among the abovementioned environmental factors that play an important role in the development of CRC, unhealthy dietary patterns are of crucial importance. In particular, a diet based on fruits and vegetables with limited consumption of red meat and confectionary is associated with a lower risk of CRC (4). In addition, there is some

evidence that food additives, even those that are officially approved, have carcinogenic properties (5). One such food additive is carrageenan, registered as E407 (refined carrageenan) and E407a (semi-refined carrageenan). Carrageenans are galactans composed of repeating disaccharides, namely β -1,3- and α -1,4linked derivatives of galactose with sulfate groups (6). Three major commercially available types of carrageenans (lambda, kappa, and iota) are widely used to improve the texture of processed meat and provide creaminess in dairy products. They are also added to food products as emulsifiers and thickeners (7). It is widely accepted and demonstrated in numerous animal studies that degraded and lowmolecular-weight carrageenans (poligeenans), officially prohibited from use as food additives, can promote ulceration and tumor development in the gut upon oral exposure (8). However, undegraded or native food-grade carrageenans have also raised carcinogenicity concerns. Nonetheless, it has been suggested that carrageenans don't act as direct carcinogens or mutagens. Instead, they stimulate the development of intestinal neoplasms via inducing inflammation, which contributes to cancer (9). Indeed, there is strong evidence that food-grade carrageenans ingested by laboratory animals (mice, rats, and guinea pigs) induce intestinal inflammation (7, 8, 10-15). Furthermore, the long-term oral consumption of carrageenans in animal experiments promotes the development of intestinal polyps, which can be precancerous (8, 16). Despite lacking mutagenic and carcinogenic properties, carrageenan creates a favorable inflammatory microenvironment for tumor development. In particular, inflammatory bowel diseases (IBD) such as Crohn's disease and ulcerative colitis can be considered an important risk factor for the development of gastrointestinal malignancies (17). Accumulating evidence indicates that consumption of carrageenans contributes to IBD pathophysiology (16, 18). Furthermore, a randomized trial showed that dietary restriction of carrageenans prevents the relapses of ulcerative Therefore, carrageenan-induced colitis (19). intestinal inflammation might be a risk factor for CRC development, and more studies are needed to elucidate both the local and systemic effects of ingested carrageenans, shedding light on the mechanisms underlying their contribution to IBD and CRC.

The purpose of our study was to evaluate the effects of orally administered E407a on the viability of lymphocytes and their cell death modes.

Materials and Methods

Animals and Groups

This research was performed using sixteen adult WAG rats of the female sex. Their weight varied from 160 to 200 g. The rats were used to form two groups on a random basis: experimental and control. Each group consisted of 8 rats. Acclimatization of the rats in the vivarium lasted for two weeks prior to the commencement of the experiment. The rats that formed the experimental group were administered 140 mg of semi-refined carrageenan per kg of weight daily for two weeks. Drinking water was used as a vehicle. The animals from the control group consumed equal volumes of drinking water with no added carrageenan.

The standards for care and housing of laboratory animals (EU Directive 2010/63/EU) were followed. The institutional ethics committee approved the study design.

Preparation of Leukocyte Suspensions

To analyze viability and cell death modes of lymphocytes in rats exposed to the E407a food additive, leukocyte suspensions were obtained from blood samples collected from rats of both groups. Initially, blood was collected in K₂EDTA Vacutainers[™] (IMPROVACUTER[®] Evacuated EDTA K, Spray Dried PET Tubes, Guangzhou, China). Then, specimens whose volume was 100 µl were transferred to 12 x 75 mm polystyrene tubes. Subsequently, 2 ml of an ammonium chloridebased lysing reagent (BD Pharmlyse[™] Lysing Buffer, lot 0070764, San Jose, USA) was added to lyse erythrocytes. Incubation with 1x Pharmlyse buffer lasted for 15 minutes at 24 °C. Solutions were centrifuged at 500g for 5 minutes. Phosphate buffered saline (PBS; pH 7.4; BDTM Cell Wash, Poland) was used to wash the leukocytes twice.

Staining Protocol

After the resuspension of cells in 1 ml of 1x Annexin-binding buffer (BD PharmingenTM Annexin V Binding Buffer, lot 8145742, BD Biosciences, San Jose, USA), 100 µl aliquots were incubated with 5 µl of fluorescein isothiocyanate (FITC)-labeled Annexin V (BD PharmingenTM FITC-Annexin V, lot 8311824, BD Biosciences, San Jose, USA) and 5 µl of 7-aminoactinomycin D (7-AAD, BD PharmingenTM, lot 8263992, BD Biosciences, San Jose, USA). After a 20-minute-long incubation, 400 µl of 1x Annexinbinding buffer was added.

Flow Cytometric Analysis

The BD FACSCanto[™] II flow cytometer (Becton Dickinson, USA) was employed for flow cytometric analysis. To collect and process the results, BD FACSDiva[™] software was used. Identification of the region of lymphocytes was based on forward versus side scatter (FSC vs. SSC). Annexin V-FITC is optimally excited at 494 nm and has a peak emission at 519 nm. Its fluorescence was registered in the standard FITC (FL1 detector) channel. 7-AAD is excited at 488 nm and emits at approximately 670 nm. Its fluorescence was detected in the FL3 channel. Annexin V and 7-AAD staining is used to identify four populations: 1 - viable cells (Annexin

V⁻, 7-AAD⁻); 2 – early apoptotic cells (Annexin V⁺, 7-AAD⁻); 3 - late apoptotic/necrotic cells (Annexin V⁺, 7-AAD⁺); and 4 - dead necrotic cells (Annexin V⁻, 7-AAD⁺).

Statistical Analysis

The Shapiro-Wilk test was used to assess the normality of the data. The outcome of the Shapiro-Wilk test substantiated the use of the non-parametric Mann-Whitney U test. Thus, non-normally distributed data were reported as median and interquartile range. Differences were considered as statistically significant if P-values were below 0.05. Numerical data were analyzed by GraphPad Prism 7.05 (GraphPad Software, USA).

Results

Annexin V/7-AAD staining is a convenient, widely recognized, and accepted approach to discriminate viable, early apoptotic, late apoptotic/necrotic, and dead necrotic cells. In this study, we assessed the way oral exposure to E407a affects the viability and cell death modes of circulating lymphocytes.

Our findings are summarized in Table 1 and Figures 1-3.

The number of viable lymphocytes that remained unstained (Annexin V⁻, 7-AAD⁻) was found to be significantly lower in rats treated with semirefined carrageenan over two weeks compared with control samples (Table 1). In addition, exposure to E407a resulted in a larger number of circulating early apoptotic cells (Annexin V⁺, 7-AAD⁻). Such cells are characterized by phosphatidylserine (PS) externalization, i.e., the translocation of PS (an anionic phospholipid located in the inner leaflet of the cell membrane in normal conditions) to the outer leaflet. Annexin V, in turn, binds to PS. At the same time, the membrane of such cells remains intact, leading to positive 7-AAD staining. The percentage of such early apoptotic lymphocytes in the experimental group of animals was revealed to be almost four-fold higher relative to the control group (Table 1). Of note, oral exposure to E407a did not affect the number of late apoptotic/ necrotic (Annexin V⁺, 7-AAD⁺) lymphocytes, i.e., cells with externalized PS and compromised cell membrane integrity. No significant difference was found between the percentages of dead necrotic (Annexin V⁻, 7-AAD⁺) circulating lymphocytes of rats untreated and treated orally with semi-refined carrageenan (Table 1).



Figure 1: An FSC/SSC dot-plot shows how the region of lymphocytes is identified.



Figure 2: A representative FL-1/FL-3 dot-plot of a control sample. Different populations of lymphocytes are demonstrated: Q1 - early apoptotic lymphocytes (Annexin V⁺, 7-AAD⁻ cells); Q2 - late apoptotic/necrotic lymphocytes (Annexin V⁺, 7-AAD⁺); Q3 - viable lymphocytes (Annexin V⁻, 7-AAD⁻ cells); and dead necrotic CD45⁺ lymphocytes (Annexin V⁻, 7-AAD⁺).



Figure 3: A representative FL-1/FL-3 dot-plot of a sample obtained from a rat exposed to E407a. Four populations of lymphocytes can be seen: Q1 - early apoptotic lymphocytes (Annexin V⁺, 7-AAD⁻ cells); Q2 - late apoptotic/necrotic lymphocytes (Annexin V⁺, 7-AAD⁺); Q3 - viable lymphocytes (Annexin V⁻, 7-AAD⁻ cells); and dead necrotic CD45⁺ lymphocytes (Annexin V⁻, 7-AAD⁺).

Discussion

Our studies performed earlier have demonstrated that carrageenan administered orally to laboratory animals promotes the development of enterocolitis, evidenced by both local and systemic alterations,

Table 1: Effects of E407a on cell death modes of circulating lymphocytes

Groups of animals	Control group	E407a intake during 14 days per	P value
Percentage of cells	(n=8), %	05	
		(n=8), %	
Viable lymphocytes (Annexin V ⁻ , 7-AAD ⁻ cells)	94.95 [94.10; 96.10]	86.90 [85.85; 88.15]	0.0006
Early apoptotic lymphocytes (Annexin V ⁺ , 7-AAD ⁻ cells)	1.45[0.88; 1.95]	5.75 [3.88; 7.95]	0.0019
Late apoptotic/necrotic lymphocytes (Annexin V ⁺ , 7-AAD ⁺ cells)	0.75 [0.10; 1.28]	1.45 [0.40; 2.90]	0.2664
Dead necrotic CD45 ⁺ lymphocytes (Annexin V ⁻ , 7-AAD ⁺ cells)	2.60 [1.83; 2.60]	3.83 [2.83; 5.10]	0.1235

including the affected morphology of both small and large bowels, changes in the blood serum cytokine profile with the elevation of circulating pro-inflammatory cytokines, oxidative stress development, and modifications of the phospholipid bilayer of cell membranes (11-13, 20). This study has supplemented conclusions concerning the toxicity of E407a. Our findings support the hypothesis that orally consumed carrageenans can result in extraintestinal effects, in particular, promotion of lymphocyte apoptosis. Our data are consistent with other studies in which food-grade refined carrageenan was reported to induce apoptosis of leukocytes (21). In addition, E407a has been shown to induce reactive oxygen species (ROS) generation by lymphocytes after oral administration (22). This suggests the possible role of ROS-mediated pathways of apoptosis induction in lymphocytes of rats exposed to this food additive. Moreover, there is some evidence that incubation of cells with carrageenans results in ROS overproduction (23), which supports the implication of ROS in lymphocyte apoptosis after the administration of E407a. However, it is interesting to note that while the direct impact of E407a on lymphocytes does not stimulate apoptosis, shortterm incubation with E407a leads to upregulation of anti-apoptotic Bcl-2 (24). Furthermore, no direct cytotoxic effects of carrageenans have been revealed in other studies (25, 26). Thus, we believe that apoptosis activation observed in our research can be mediated by indirect mechanisms via an inflammatory response that emerges locally in the intestine. A plethora of animal studies that support the toxicity of E407 and E407a and the controversial data of experiments performed on cell cultures suggest that the effects of carrageenans in the body are complex and involve combined interactions. In particular, interactions of carrageenan with the gut microbiota may mediate the pro-inflammatory response to the food additive. This hypothesis is in agreement with numerous findings that have revealed the role of carrageenan in enhancing bacterial

lipopolysaccharide (LPS)-induced upregulation of pro-inflammatory cytokines (27, 28). Furthermore, carrageenan has been shown to worsen bacterial intestinal inflammation in mice. Authors state that κ -carrageenan stimulates the Bcl10-NF- κ B-mediated pathway and thereby activates LPS-induced secretion of IL-8 while promoting the expression of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-kB). This transcriptional factor is known to be highly pro-inflammatory (28).

Our study sheds light on the effects of E407a consumption on cell death modes of lymphocytes. However, more well-designed research is needed to study the mechanisms involved in carrageenaninduced activation of apoptosis. In addition, this research reinforces the need for novel studies, primarily clinical ones, to evaluate the role of carrageenans in the development of IBD and CRC.

Conclusion

Our data indicate that the food additive E407a stimulates apoptosis of circulating lymphocytes.

Funding

This study was performed as a fragment of research entitled Biochemical Mechanisms for the Induction of Intestinal Inflammation and Ways of its Correction (state registration number 0120U102645).

Financial Disclosure: The authors state that no financial interests are involved.

Authors Contributions: Study concept and design: Tkachenko. Data analysis and interpretation: Tkachenko and Onishchenko. Drafting: Tkachenko. Critical revision of the manuscript for important intellectual content: Tkachenko and Onishchenko. Statistical analysis: Tkachenko.

Conflicts of interests: None declared.

References

- 1. Golshani G, Zhang Y. Advances in immunotherapy for colorectal cancer: a review. Therapeutic Advances in Gastroenterology. January 2020. doi:10.1177/1756284820917527
- 2. Alexander PG, McMillan DC, Park JH. The local inflammatory response in colorectal cancer - Type, location or density? A systematic review and meta-analysis. Cancer Treat Rev. 2020;83:101949.
- 3. Kuipers EJ, Grady WM, Lieberman D, Seufferlein T, Sung JJ, Boelens PG, et al. Colorectal cancer. Nat Rev Dis Primers. 2015;1:15065. doi: 10.1038/ nrdp.2015.65.
- 4. Tabung FK, Brown LS, Fung TT.

Dietary patterns and colorectal cancer risk: a review of 17 years of evidence (2000-2016). Curr Colorectal Cancer Rep. 2017;13(6):440-454. doi:10.1007/ s11888-017-0390-5

- Gultekin F, Yasar S, Gurbuz N, Ceyhan BM. Food additives of public concern for their carcinogenicty. J. Nutr. Health Food Sci. 2015;3:1-6
- 6. Yermak IM, Mischchenko NP, Davydova VN, Glazunov VP, Tarbeeva DV, Kravchenko AO, et al. Carrageenans-sulfated polysaccharides from red seaweeds as matrices for the inclusion of Echinochrome. Mar Drugs. 2017;15(11):337. doi: 10.3390/

md15110337.

- Necas J, Bartosikova L. Carrageenan: a review. Veterinarni Medicina 2013;58:187-205.
- Tobacman JK. Review of harmful gastrointestinal effects of carrageenan in animal experiments. Environ Health Perspect. 2001;109(10):983– 994. doi:10.1289/ehp.01109983
- 9. Heikenwälder H, Heikenwälder M. Krebs - Lifestyle und Umweltfaktoren als Risiko. Bindemittel, Ballaststoffe und Darmentzündungen. Springer; 2019. P. 69-78.
- David S, Shani Levi C, Fahoum L, Ungar Y, Meyron-Holtz EG, Shpigelman A, et al. Revisiting the

carrageenan controversy: do we really understand the digestive fate and safety of carrageenan in our foods? Food Funct. 2018;9(3):1344-1352. doi: 10.1039/c7fo01721a.

- 11. Tkachenko AS, Onishchenko AI, Gorbach TV, Gubina-Vakulyck GI. O-6-methylguanine-DNA methyltransferase (MGMT) overexpression in small intestinal mucosa in experimental carrageenaninduced enteritis. Malay. J. Biochem. Mol. Biol. 2018;21(3):77-80.
- Tkachenko A, Marakushyn D, Kalashnyk I, Korniyenko Y, Onishchenko A, Gorbach T, et al. A study of enterocyte membranes during activation of apoptotic processes in chronic carrageenaninduced gastroenterocolitis. Med Glas (Zenica). 2018;15(2):87-92. doi: 10.17392/946-18.
- 13. Gubina-Vakyulyk GI, Gorbach TV, Tkachenko AS, Tkachenko MO. Damage and regeneration of small intestinal enterocytes under the influence of carrageenan induces chronic enteritis. Comp. Clin. Path. 2015;24(6):1473–1477. https://doi. org/10.1007/s00580-015-2102-3
- 14. Bhattacharyya S, Xue L, Devkota S, Chang E, Morris S, Tobacman JK. Carrageenan-induced colonic inflammation is reduced in Bcl10 null mice and increased in IL-10deficient mice. Mediators Inflamm. 2013;2013(10):397642.
- 15. Bhattacharyya S, Gill R, Chen M-L, Zhang F, Linhardt RJ, Dudeja PK, et al. Toll-like receptor-4 mediates induction of Bcl10-NFκB-IL-8 inflammatory pathway by carrageenan in human intestinal epithelial cells. J Biol Chem. 2008;16(283):10550–8
- **16.** Martino JV, Van Limbergen J, Cahill LE. The role of carrageenan and carboxymethylcellulose in

the development of intestinal inflammation. Front Pediatr. 2017;5:96. doi: 10.3389/fped.2017.00096.

- Biancone L, Armuzzi A, Scribano ML, Castiglione F, D'Incà R, Orlando A, et al. Cancer risk in inflammatory bowel disease: a 6-year prospective multicenter nested case-control IG-IBD study. Inflamm Bowel Dis. 2020;26(3):450-459. doi: 10.1093/ibd/ izz155.
- **18.** Rizzello F, Spisni E, Giovanardi E, Imbesi V, Salice M, Alvisi P, et al. Implications of the westernized diet in the onset and progression of IBD. Nutrients 2019;11:1033.
- 19. Bhattacharyya S, Shumard T, Xie H, Dodda A, Varady KA, Feferman L, et al. A randomized trial of the effects of the no-carrageenan diet on ulcerative colitis disease activity. Nutr Healthy Aging. 2017;4(2):181-192. doi: 10.3233/NHA-170023.
- 20. Tkachenko AS, Gubina-Vakulyck GI, Klochkov VK, Kavok NS, Onishchenko AI, Gorbach TV, et al. Experimental evaluation of the impact of gadolinium orthovanadate GdVO₄:Eu³⁺ nanoparticles on the carrageenan-induced intestinal inflammation. Acta Medica (Hradec Králové). 2020;63(1):18–24 https://doi. org/10.14712/18059694.2020.11
- Kopanytsia OM, Marushchak MI, Krynytska IY. Carrageenan induces cell death in rats blood. International Journal of Medicine and Medical Research. 2018;4(1):67-70. https://doi.org/10.11603/ ijmmr.2413-6077.2018.1.8979
- Tkachenko A. Reactive oxygen species (ROS) generation by lymphocytes in rats treated with a common food additive E407a. J Clin Med Kaz. 2020;1(55):22-26. DOI: 10.23950/1812-2892-JCMK-00744
 Sokolova EV, Karetin Y, Davydova

VN, Byankina AO, Kalitnik AA, Bogdanovich LN, et al. Carrageenans effect on neutrophils alone and in combination with LPS in vitro. J Biomed Mater Res A. 2016;104(7):1603-9. doi: 10.1002/ jbm.a.35693.

- 24. Tkachenko AS, Onishchenko AI, Lesovoy VN, Myasoedov VV. Common food additive E407a affects BCL-2 expression in lymphocytes in vitro. Studia Univ. VG, SSV, 2019;29(4):169-76.
- 25. McKim JM, Willoughby JA Sr, Blakemore WR, Weiner ML. Clarifying the confusion between poligeenan, degraded carrageenan, and carrageenan: A review of the chemistry, nomenclature, and in vivo toxicology by the oral route. Crit Rev Food Sci Nutr. 2019;59(19):3054-3073. doi: 10.1080/10408398.2018.1481822.
- 26. McKim JM Jr, Baas H, Rice GP, Willoughby JA Sr, Weiner ML, Blakemore W. Effects of carrageenan on cell permeability, cytotoxicity, and cytokine gene expression in human intestinal and hepatic cell lines. Food Chem Toxicol. 2016;96:1-10. doi: 10.1016/j.fct.2016.07.006.
- 27. Wu W, Zhen Z, Niu T, Zhu X, Gao Y, Yan J, et al. κ-Carrageenan enhances lipopolysaccharideinduced interleukin-8 secretion by stimulating the Bcl10-NF-κB Pathway in HT-29 cells and aggravates C. freundii-induced inflammation in mice. Mediators Inflamm. 2017;2017:8634865. doi: 10.1155/2017/8634865.
- 28. Ogata M, Matsui T, Kita T, Shigematsu A. Carrageenan primes leukocytes to enhance lipopolysaccharideinduced tumor necrosis factor alpha production. Infect Immun. 1999;67(7):3284–3289.