

Cell organelle-shaped liposomes: A novel approach to present the stable intracellular drug delivery systems

Amir Azadi^{1, 2}, Hajar Ashrafi^{3*}

Abstract

Caveolae are lipid raft-enriched flask-shaped, expose in the plasma membrane of various cell types. It has become clear now that caveolae and their caveolin "marker proteins" are associated in a several cellular procedures including endocytosis, lipid homeostasis, signal transduction, and tumorigenesis. Caveolin has been shown to have high binding affinity for cholesterol and sphingolipids. Caveolin oligomers construct filamentous networks that are believed to stabilize the membrane. Liposomes are the well-known drug delivery systems with spherical shape that can be produced from natural non-toxic phospholipids and cholesterol. Liposomes have been used as a considerable tool in biology, biochemistry, medicine, and drug delivery. The utilization of liposomes as a drug-delivery system has become more attractive in carrying systemically administered drugs with narrow therapeutic windows. The similarity between plasma membrane and liposomes from several points of view gives hope that the incorporation of caveolin in the phospholipid bilayer structures of liposomes can result in tightening and therefore stabilizing and long circulation of these structures.

Keywords: Caveolin, Liposome, Long-circulation, Phospholipid bilayer tightening.

1. Introduction

Caveolae are the small invaginations of the plasma membrane with the size of 50 to 100 nm (1). A long with the physiologic-based influential roles of these important structures, it has become clear now that caveolae and their caveolin "marker proteins" are associated in a several cellular processes including lipid homeostasis, endocytosis, signal transduction, and tumorigenesis (2) (Figure 1). The caveolin family is consisted of three definite proteins including caveolin-1, caveolin-2, and caveolin 3 (Cav-1, -2,-3) (3, 4). The caveolin proteins have a variety features which are important not only for selective localization to caveolae but also for compelling the invagination of these com-

••••••••••••

Corresponding Author: Hajar Ashrafi, Department of Pharmaceutics, School of Pharmacy, International branch, Shiraz University of Medical Sciences Shiraz, Iran.
Email: hashrafi@sums.ac.ir

plexes. Cav-1 has been demonstrated to have high binding affinity for cholesterol and sphingolipids (5, 6). This property, as well as three carboxyterminal lipid modifications (palmitoylations), stabilizes and targets Cav-1 to caveolae (7). Cav-1 oligomers form filamentous networks that are believed to stabilize the membrane and define the size and shape of caveolae (8). Caveolae can be divided from cells as cholesterol and sphingomyelin-enriched microdomains that are traditionally found decorating both apical and/or basolateral membranes in terminally differentiated cells (9). Identification and cloning of a caveolae coat-associated protein, Cav-1 is also possible (10, 11).

Liposomes and their characterizations

Liposomes are the well-known drug delivery systems with spherical shape that can be pro-

¹Pharmaceutical Sciences Research Center, Shiraz University of Medical Sciences, Shiraz, Iran.

²Department of Pharmaceutics, School of Pharmacy, Shiraz University of Medical Sciences, Shiraz, Iran.

³Department of Pharmaceutics, School of Pharmacy, International branch, Shiraz University of Medical Sciences, Shiraz, Iran

Amir Azadi et al.

duced from natural non-toxic phospholipids and cholesterol. Phospholipids, triglycerides, and cholesterol are the main constituents of liposomes. In fact, liposomes are phospholipid bilayers with an entangled aqueous extent. The first suggested utilized of liposomes came from Weismann et al. in 1969 (12). Since then, liposomes have been used as a considerable tool in biology, biochemistry, medicine, and drug delivery (13). Because of their wide size range, hydrophobic/hydrophilic properties and, demonstrated biocompatibility, liposomes are considered as a hopeful system for drug delivery. Furthermore, drugs with different lipophilicities can be encapsulated into liposomes (14). The utilization of these lipid-based vesicles as a drugdelivery system has become more attractive over the last decades, because of their biocompatibility and flexibility in carrying systemically administered drugs such as chemotherapeutics and antibiotics with restricted therapeutic windows. Liposomes can be conjugated to antibodies or ligands to provide target-specific drug delivery possibility. They are classified into three categories based on their size and number of bilayers. Small unilamellar vesicles (SUV) are surrounded by a single lipid bilayer and are 25–50 nm in diameter. Large unilamellar vesicles (LUV) are a heterogeneous group of vesicles similar to SUVs but larger in size, while enclosed by a single lipid bilayer, too. Multilamellar vesicles (MLV); however, consist of several lipid bilayers separated from one another by a layer of aqueous solution (15). Moreover, liposomes are widely used as carriers for small drugs and macromolecules in drug delivery (16).

Hypothesis: Caveolin can act as a stabilizing and long circulating agent in liposomes

The similarity between plasma membrane and liposomes from the view point of lipid combination, bilayer structure, and the possibility of the insertion of various ligands in these structures, along with high binding affinity of Cav-1 for cholesterol is an optimistic sign that the incorporation of Cav-1 in the phospholipid bilayer structures of liposomes-similar to what is naturally observed in the lipid rafts of plasma membrane — can result in

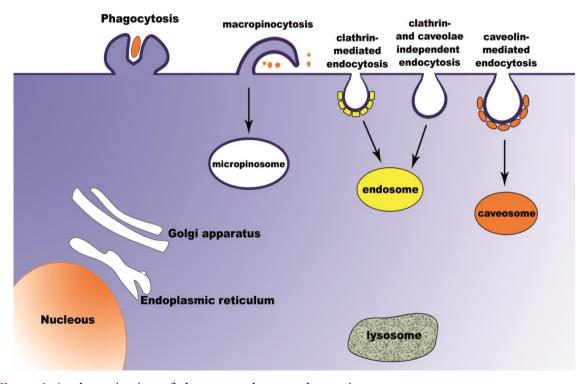


Figure 1. A schematic view of plasma membrane endocytosis processes.

tightening and therefore stabilizing of these structures. This tight and stabilized bilayer can decrease insertion of plasma proteins and also reduce reticuloendothelial system (RES) clearance of the liposomes. The stabilized liposomes can passively accumulate inside other tissues and/or organs. This aspect, named passive targeting, is especially apparent in solid tumors enduring angiogenesis. The existence of a disconnected endothelial lining in the tumor vasculature during angiogenesis promotes extravasation of liposomal formulations into the interstitial space, where they accumulate due to the lack of efficient lymphatic effluent of the tumor, and function as a controlled drug-release system. This causes the preferential accumulation of liposomes in the tumor area (a process known as enhanced permeation and retention effect EPR). Therefore, this type of liposomal drug delivery system can especially be appropriate for chemotherapeutic agent delivery and targeting.

Incorporation of the caveolin can be accomplished during liposome formation by detergent dialysis or post liposome formation by conjugation to reactive lipids. Such strategies are usual for the incorporation of the targeting moieties (13,

17). In addition to stabilizing-which in turn causes long circulation-and its consequences, Cav-1 may, this time, play its routine role of signal transduction for liposomes instead of natural cells, the revolutionary effects of which goes without saying.

Concluding remarks

The most significant properties of caveolin-modified liposomes can strongly be prolonged blood circulation and thus improved distribution in perfused tissues. Taking into account these considerations and the great advantages of caveolin-modified liposomes in decreasing specific drug toxicity and in passively targeting the incorporated molecules to the site of action, new and "improved" liposomal formulations designed for different therapeutic and diagnostic areas may be expected to arrive on the pharmaceutical research.

Acknowledgments

The authors gratefully acknowledge *Dr. M. Rahimi* and *Mr. J. Ebrahimi* for their valuable contributions.

Conflict of Interest

None declared.

References

- 1. Cheng JPX, Nichols BJ. Caveolae: One Function or Many? *Trends Cell Biol*. 2016;26:177-89.
- 2. Moore DH, Ruska H. The fine structure of capillaries and small arteries. *J Biophys Biochem Cytol*. 1957;3:457-62.
- 3. Glenney JR. The sequence of human caveolin reveals identity with VIP21, a component of transport vesicles. *FEBS Lett.* 1992;314:45-8.
- 4. Scherer PE, Okamoto T, Chun M, Nishimoto I, Lodish HF, Lisanti MP. Identification, sequence, and expression of caveolin-2 defines a caveolin gene family. *Proc Natl Acad Sci U S A*. 1996;93:131-5.
- 5. Fra AM, Masserini M, Palestini P, Sonnino S, Simons K. A photo-reactive derivative of ganglioside GM1 specifically cross-links VIP21-caveolin on the cell surface. *FEBS Lett.* 1995;375:11-4.
- 6. Thiele C, Hannah MJ, Fahrenholz F, Huttner WB. Cholesterol binds to synaptophysin and is required for biogenesis of synaptic vesicles. *Nat Cell Biol.* 2000;2:42-9.

- 7. Dietzen DJ, Hastings WR, Lublin DM. Caveolin is palmitoylated on multiple cysteine residues Palmitoylation is not necessary for localization of caveolin to caveolae. *J Biol Chem.* 1995;270:6838-42.

 8. Pelkmans L, Bürli T, Zerial M, Helenius A.
- Caveolin-stabilized membrane domains as multifunctional transport and sorting devices in endocytic membrane traffic. *Cell.* 2004;118:767-80.
- 9. Razani B, Woodman SE, Lisanti MP. Caveolae: from cell biology to animal physiology. *Pharmacol Rev.* 2002;54:431-67.
- 10. Glenney JR, Soppet D. Sequence and expression of caveolin, a protein component of caveolae plasma membrane domains phosphorylated on tyrosine in *Rous sarcoma* virus-transformed fibroblasts. *Proc Natl Acad Sci USA*. 1992;89:10517-21. 11. Rothberg KG, Heuser JE, Donzell WC, Ying
- 11. Rothberg KG, Heuser JE, Donzell WC, Ying Y-S, Glenney JR, Anderson RGW. Caveolin, a protein component of caveolae membrane coats. *Cell.* 1992;68:673-82.
- 12. Bangham AD, Standish MM, Watkins JC. Diffusion of univalent ions across the lamellae of

- swollen phospholipids. *J Mol Biol.* 1965;13:238-IN27.
- 13. Schiffelers RM, Koning GA, ten Hagen TLM, Fens MHAM, Schraa AJ, Janssen APCA, et al. Anti-tumor efficacy of tumor vasculature-targeted liposomal doxorubicin. *J Control Release*. 2003;91:115-22.
- 14. Kulkarni PR, Yadav JD, Vaidya KA. Liposomes: a novel drug delivery system. *Int J Curr Pharm Res.* 2011;3:10-8.
- 15. Akbarzadeh A, Rezaei-Sadabady R, Davaran S, Joo SW, Zarghami N, Hanifehpour Y, *et al.* Liposome: classification, preparation, and applications. *Nanoscale Res Lett.* 2013;8:102.
- 16. Sharma S, Mishra L, Grover I, Gupta A, Kaur K. Liposomes: vesicular system an overview. *Int J Pharm Pharm Sci.* 2010;2:11-7.
- 17. Hood JD, Bednarski M, Frausto R, Guccione S, Reisfeld RA, Xiang R, et al. Tumor regression by targeted gene delivery to the neovasculature. *Science*. 2002;296:2404-7.